Table 1 Inhibition of histamine release from rat mast cells

			$ID_{50} \mu g/mI$		
Histamine liberator	(μ <i>g/ml)</i>	Indomethacin	Meclofenamate	PPP	PGE_2
Compound 48/80	(0.16)	21.5	3.5	0.7	16.0
Phospholipase-A	(5.0)	13.5	2.0		_
Adenosine triphosphate	(200)	10.0	3.0	2.5	7.0
Antigen-egg albumen	(1000)	13.0	6.5	12.0	10.5

The concentration of inhibitor giving 50% reduction of histamine release (ID_{50}), following 5 min pre-incubation with mast cells, was determined from at least 5 experiments. Control histamine release, with the concentration of each liberator shown, was 30–40% of total histamine content of the cells.

was 0.5 µg/ml). In contrast, the anti-inflammatory steroid, betamethasone (100 µg/ml), only slightly inhibited (18-25%) antigen- or 48/80-induced histamine release. Polyphloretin phosphate also reduced histamine release (Table 1) but was more potent than di-4-phloretin phosphate (ID₅₀ against 48/80-induced release was 0.7 and 3.3 μg/ml respectively), unlike the relative potency of these compounds as either prostaglandin antagonists or as inhibitors of prostaglandin inactivation (Crutchley & Piper, 1974). Prostaglandin E₂ (PGE₂) reduced histamine release stimulated by the various liberators shown (Table 1) but exhibited a shallow doseinhibition relationship; PGE₂ or its (15S)-15 methyl analogue (1 µg/ml) inhibited antigen-induced release by 20-30%.

These observations that non-steroid antiinflammatory drugs inhibit histamine release supports the finding that indomethacin reduces mast cell degranulation (Taylor, Francis, Sheldon & Roitt, 1974). The mechanism underlying this inhibitory activity is not yet known, but may involve effects on cyclic AMP levels via phosphodiesterase inhibition, on membrane stability or on calcium-ion mobilization. Thus, assessment of a role for endogenous prostaglandins in mast cells is made difficult by these possible actions of aspirin-like drugs. However, the ability of prostaglandins to reduce mast cell histamine release could reflect a patho-physiological modulator role during anaphylaxis. This work was supported by a grant from the M.R.C. to Professor G.P. Lewis. Prostaglandins were kindly supplied by the Upjohn Company, Kalamazoo.

References

- CRUTCHLEY, D.J. & PIPER, PRISCILLA J. (1974). Prostaglandin inactivation by guinea-pig lung and its inhibition. Br. J. Pharmac., 52, 197-203.
- FOREMAN, J.C., MONGAR, J.L. & GOMPERTS, B.D. (1973). Calcium ionophores and movement of calcium ions following the physiological stimulus to a secretory process. *Nature (Lond.)*, 245, 249–251.
- KALINER, M. & AUSTEN, K.F. (1974). Cyclic AMP, ATP and reversed anaphylactic histamine release from rat mast cells. J. Immun., 112, 664-672.
- SHORE, P.A., BURKHALTER, A. & COHN, V.H. (1959). A method for the fluorometric assay of histamine in tissues. J. Pharmac. exp. Ther., 127, 182–186.
- TAYLOR, W.A., FRANCIS, D.H., SHELDON, D. & ROITT, I.M. (1974). Anti-allergic actions of disodium cromoglycate and other drugs known to inhibit cyclic 3',5'-nucleotide phosphodiesterase. *Int. Archs. Allergy appl. Immun.*, 47, 175–193.
- VANE, J.R. (1971). Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature*, *New Biol.*, 231, 232-235.

Steroid hormone receptors in brain and pituitary

M. GINSBURG & P.J. THOMAS

Department of Pharmacology, Chelsea College, University of London.